



Review

A review on the role of ZEB1-AS1 in human disorders



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ABSTRACT

ZEB1 Antisense RNA 1 (ZEB1-AS1) is a type of RNA characterized as long non-coding RNA (lncRNA). This lncRNA has important regulatory roles on its related gene, Zinc Finger E-Box Binding Homeobox 1 (ZEB1). In addition, role of ZEB1-AS1 has been approved in diverse malignancies such as colorectal cancer, breast cancer, glioma, hepatocellular carcinoma and gastric cancer. ZEB1-AS1 serves as a sponge for a number of microRNAs, namely miR-577, miR-335-5p, miR-101, miR-505-3p, miR-455-3p, miR-205, miR-23a, miR-365a-3p, miR-302b, miR-299-3p, miR-133a-3p, miR-200a, miR-200c, miR-342-3p, miR-214, miR-149-3p and miR-1224-5p. In addition to malignant conditions, ZEB1-AS1 has functional role in non-malignant conditions like diabetic nephropathy, diabetic lung, atherosclerosis, Chlamydia trachomatis infection, pulmonary fibrosis and ischemic stroke. This review outlines different molecular mechanisms of ZEB1-AS1 in a variety of disorders and highlights its importance in their pathogenesis.

1. Introduction

After a long time of debate on whether long non-coding RNAs (lncRNAs) should be considered as junk DNA or functional ones, it is now concluded that these molecules have important biological characteristics [12,13,47,51]. In general, an lncRNA is a type of RNA that is not translated to protein, is longer than 200 bp and is usually transcribed by RNA POL II [14]. Although they are called “non-coding”, recent investigations have proved that a number of lncRNAs can actually encode proteins or small peptides [4]. Regarding their critical role in the pathogenesis of both malignant and non-malignant disorders, uprising studies are being published every day to emphasize on the function of lncRNAs, and how these molecules can revolutionize therapeutic approaches for treatment of different conditions.

ZEB1 Antisense RNA 1 (ZEB1-AS1, also known as HSNLNG0077083 in LNCipedia database), is a classified lncRNA. ZEB1-AS1 usually resides in nucleus and according to NCBI, its significant expression has been verified in endometrium, thyroid and ovary. Its genomic position is on the short arm

of chromosome 10 (10p11.22) (Fig. 1). According to ensembl genome browser, alternative splicing contributes to 12 different transcripts of ZEB1-AS1 (https://asia.ensembl.org/Homo_sapiens/Gene/Summary?db=core;g=ENSG00000237036;r=10:31206278-31320447). Different studies have verified that ZEB1-AS1 has a positive correlation with ZEB1 via epigenetics modifications and induces its stability [36,54]. Altogether, ZEB1-AS1 has been shown to have oncogenic roles in different cancers such as glioma, non-small cell lung cancer, colorectal, gastric and prostate cancer. In addition to malignant conditions, ZEB1-AS1 plays a central position in the pathogenesis of non-malignant disorder like atherosclerosis, ischemic stroke and even Chlamydia trachomatis infection. Several studies also correlate levels of ZEB1-AS1 to diabetes complications, including diabetic nephropathy. In this review, we aim to pinpoint molecular mechanisms of ZEB1-AS1 in different disorders and highlight its importance in pathogenesis.

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2. Role of ZEB1-AS1 in cancers

2.1. In vitro studies

Amongst different types of cancer, functional role of ZEB1-AS1 has been studied to a greater extent in colorectal cancer (CRC). In the case of drug sensitivity, it has been shown that knockdown of ZEB1-AS1 in CRC cell lines HT-29 and HCT116 increases 5-fluorouracil sensitivity [28]. Two major signaling pathways in CRC are affected by ZEB1-AS1. In SW480 and HCT116 cell lines, downregulation of ZEB1-AS1 inhibits PI3K/AKT signaling pathway and contributes to decreased colony formation and epithelial-mesenchymal transition (EMT) [57]. In another study conducted by Lv S.Y et al., ZEB1-AS1 can positively affect β -catenin and TCF4 expression, so if downregulated, Wnt/ β -catenin signaling pathway is attenuated and apoptosis is induced [38]. One of the main regulatory mechanisms of lncRNAs is molecular sponging [46]. In SW480, LOVO, HT29 and PKO cell lines, ZEB1-AS1 acts as a molecular sponge for miR-141-3p. After knocking down ZEB1-AS1, miR-141-3p levels rises and inhibits proliferation of cancer cells [66].

Similar to CRC, ZEB1-AS1 has an oncogenic role in hepatocellular carcinoma (HCC) and its downregulation is in favor of benign features. Interestingly, ZEB1-AS1 can regulate levels of both E2F1 and E2F2 in HCC [26,44]. With regards to E2F1 regulation, miR-299-3p/E2F1 axis

has been approved as a functional axis. ZEB1-AS1 sponges miR-299-3p, and this microRNA targets 3'-UTR of E2F1, so E2F1 levels rises [44]. Considering transforming roles of E2F1 [9], mentioned axis contributes to malignant features like increased proliferation, migration and invasiveness in HCC cells [44]. E2F2 is another transcription factor acting a promoter or inhibitor of proliferation [8]. In HCC cell lines HepG2, 293 T and HCCLM6, ZEB1-AS1 downregulation contributes to upregulation of miR-365a-3p and subsequent targeting of E2F2, ultimately resulting in favorable (normal-like) properties of cancer cells [26].

Zinc Finger E-Box Binding Homeobox 1 (ZEB1) is known as master regulator of EMT and its dysregulation contributes to different types of cancer [45,56]. Identical to many lncRNAs, ZEB1-AS1 can regulate expression of its related gene ZEB1 in a positive manner. For instance, in glioma cell lines U87 and U251, interference of ZEB1-AS1 contributes to reduction of ZEB1 via miR-200c/141 and as expected, reduced levels of ZEB1 is associated with absence of malignant properties (Fig. 2) [42]. Relation of ZEB1-AS1 and ZEB1 is also explicated in pancreatic cancer cell lines, in which depletion of HIF-1 α is followed by reduction of ZEB1-AS1 and ZEB1 expression [23].

One of the exact molecular mechanisms in relation to ZEB1-AS1/ZEB1 axis was demonstrated by Su W et al. in DU145 and PC3 prostate cancer cells [55]. Compellingly, ZEB1-AS1 can recruit MLL1 to the promoter region of ZEB1, induce H3K4me3 modifications and thereby

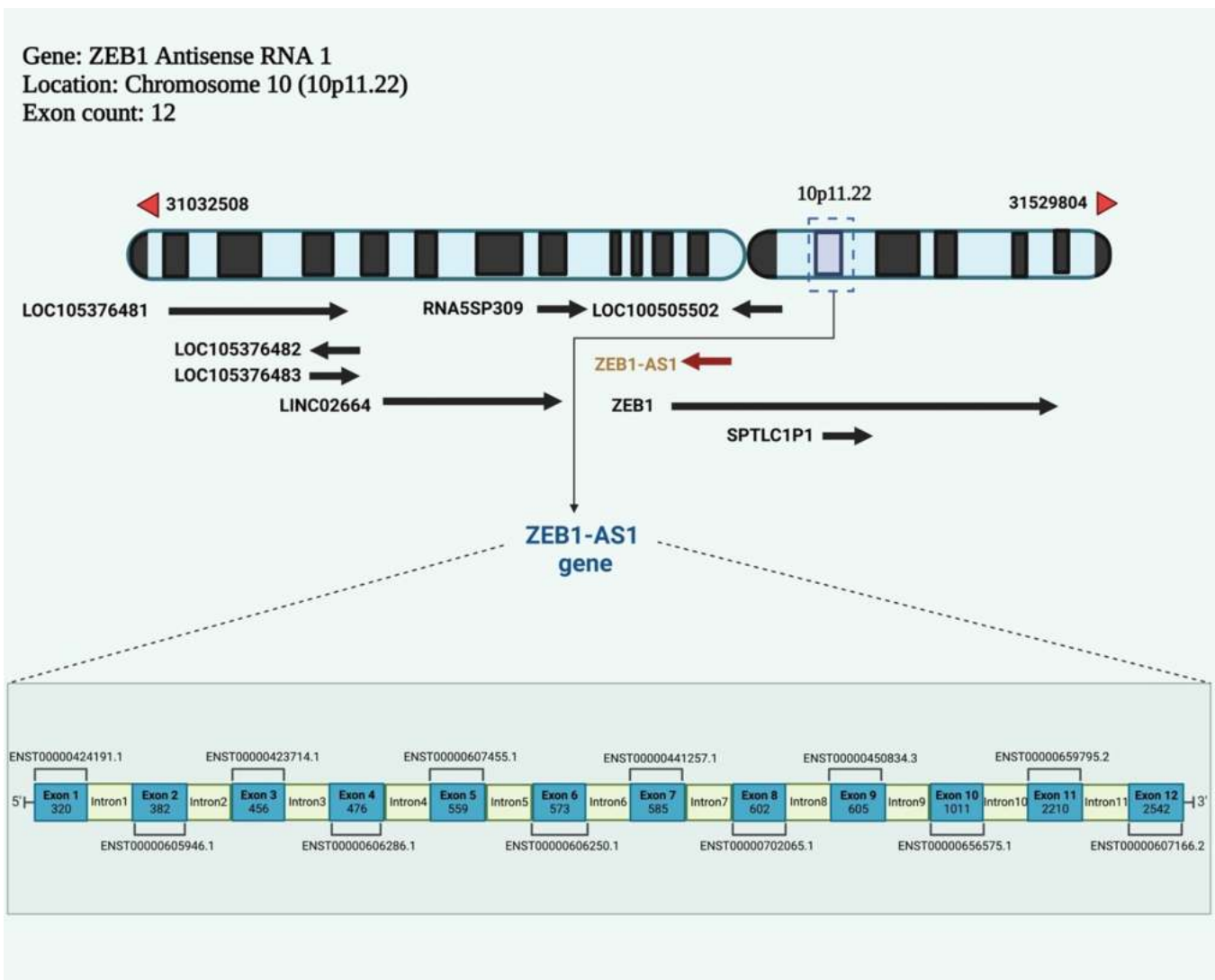


Fig. 1. Illustrates newly discovered lncRNA ZEB1-AS1 has a coding gene with 12 exons and located on short arm of chromosome 10 (10p11.22).

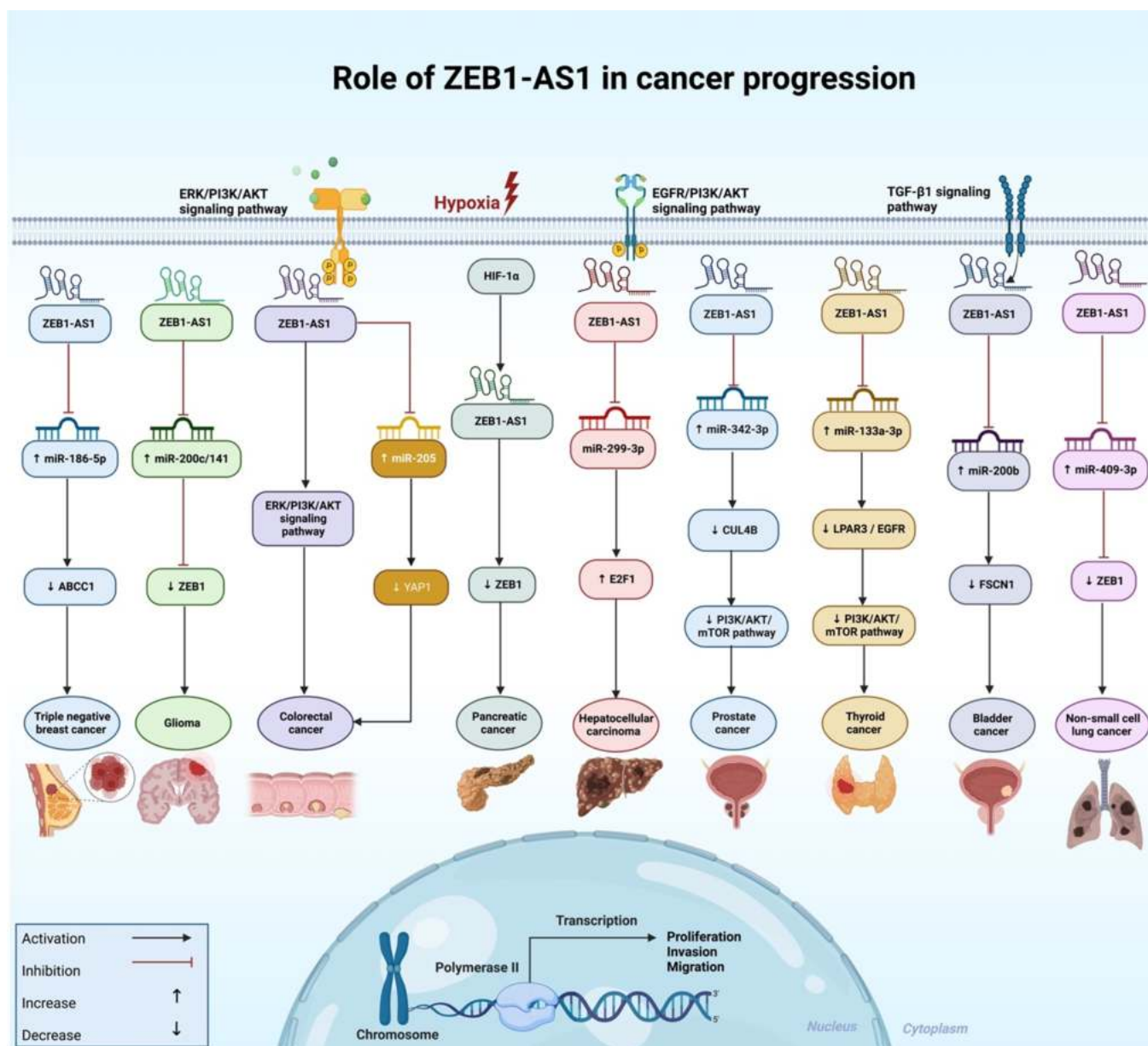


Fig. 2. The progression of ZEB1-AS1 in various tumors is depicted, which promotes tumor cell proliferation, invasion, migration, and survival by targeting certain genes and sponging distinct types of miRNAs.

inducing ZEB1. After forced inhibition of ZEB1-AS1, reduced level of ZEB1 is observed and this phenomenon is intertwined with reduced proliferation and migration in prostate cancer [55].

Without a doubt, ZEB1-AS1 acts as a transforming lncRNA and downregulation of it could be beneficial and in favor of benign properties. Table 1 summarizes functional studies on impact of ZEB1-AS1 in different types of cancer cell line.

2.2. In vivo studies

Analogous to in vitro assays, ZEB1-AS1 has been shown to be a transforming lncRNA in xenograft models, specifically mice models. Two different studies in non-small cell lung cancer have supported this view. After knocking down ZEB1-AS1, tumor growth and weight reduces in comparison to control group in BALB/c nude mice [22,25].

In intrahepatic cholangiocarcinoma, after subcutaneous injection of HuCCT1 cells in which ZEB1-AS1 was knocked down, distant metastasis

to lung was inhibited, recommending an anti-metastatic role for this lncRNA in vivo [21]. This phenomenon has also been affirmed in pancreatic cancer [23], oral squamous cell carcinoma [62] CRC [15] and HCC [41].

Cumulatively, different in vivo studies in different malignancies suggest an oncogenic role for ZEB1-AS1. Table 2 summarizes functional role of ZEB1-AS1 in vivo.

2.3. Studies in clinical samples

Different studies in a wide variety of clinical samples have proved the oncogenic roles of ZEB1-AS1 (Table 3). An *in silico* analysis of RNA transcriptome dataset of TCGA and related clinical characteristics of CRC patients obtained from the USCS Xena website has revealed that ZEB1-AS1 is among m6A-related prognostic lncRNAs that confer risk of CRC [28]. Expression of this lncRNA has been higher in high risk group of CRC compared with low risk group [28]. Moreover, expression of

Table 1

Role of ZEB1-AS1 in cancer cell lines (Δ : knock-down or deletion, \uparrow : overexpression, \rightarrow : results in, EMT: Epithelial-mesenchymal transition, DOX: doxorubicin, UA: Ursolic acid, 5-FU: 5-fluorouracil, DDP: cisplatin, PTX: paclitaxel).

| Tumor types | Targets/Regulators and signaling pathways | Cell lines | Functions | Ref |
|-------------------------------------|---|---|--|------|
| Triple negative breast cancer | miR-186-5p/ ABCC1 axis | DOX-resistant MDA-MB-468 and MDA-MB-436 | Treatment with UA: \downarrow ZEB1-AS1 \rightarrow \uparrow miR-186-5p \rightarrow \downarrow ABCC1: \downarrow proliferation \uparrow sensitivity to DOX | [34] |
| | ZEB1/ELAVL1 | MDA-MB-436, MDA-MB-453, MCF-7 and MDA-MB-231 | Δ ZEB1-AS1 (which binds to ELAVL1) \rightarrow \downarrow ZEB1: \downarrow proliferation \uparrow apoptosis | [36] |
| Glioblastoma | - | U87MG | Δ ZEB1-AS1: \downarrow proliferation \downarrow migration \uparrow apoptosis | [74] |
| Glioma | miR-200c/141/ZEB1 axis | U87, U251, LN18, U118, and T98G | Δ ZEB1-AS1 \rightarrow \uparrow miR-200c/141 \rightarrow \downarrow ZEB1: \downarrow proliferation \downarrow migration \downarrow colony formation \downarrow motility | [42] |
| | miR-577 | A172, U87, T98G and SHG44 | Δ ZEB1-AS1 (which acts as a molecular sponge) \rightarrow \uparrow miR-577: \downarrow proliferation \downarrow migration \downarrow invasion | [65] |
| | ZEB1 | HS683, T98G, U87, and U251 | Δ ZEB1-AS1 \rightarrow \downarrow ZEB1: \downarrow proliferation \downarrow migration \downarrow invasion \uparrow apoptosis | [37] |
| Colorectal cancer | - | HT-29 and HCT116 | Δ ZEB1-AS1 \downarrow proliferation \downarrow 5-FU resistance \downarrow EMT process | [28] |
| | - | Caco-2, HT-29, HCT116 | Up-regulated ZEB1-AS1 is associated with \uparrow sensitivity to: nelarabine, palbociclib, fluphenazine, asparaginase, LEE-011, Ifosfamide, hydroxyurea and dexrazoxane | [6] |
| | ERK/PI3K/AKT | SW480 and HCT116 | Δ ZEB1-AS1 \rightarrow \downarrow ERK/PI3K/AKT \downarrow colony formation \downarrow EMT process | [57] |
| | miR-335-5p/APOC1 axis | HCT116 | Δ ZEB1-AS1 \rightarrow \uparrow miR-335-5p \rightarrow \downarrow APOC1: \downarrow invasion \downarrow migration | [33] |
| | miR-141-3p | SW480, LOVO, HT29 and PKO | Δ ZEB1-AS1 (which acts as a molecular sponge) \rightarrow \uparrow miR-141-3p: \downarrow proliferation \uparrow cell cycle arrest | [66] |
| | miR-101/ZEB1 axis | SW480, DLD-1, HCT116, SW620, and HT29 | Δ ZEB1-AS1 (which acts as a molecular sponge) \rightarrow \uparrow miR-101 \rightarrow \downarrow ZEB1: \downarrow proliferation \downarrow migration | [69] |
| | p15 | SW480, DLD-1, HCT116, SW620 | Δ ZEB1-AS1 \rightarrow \uparrow p15 \downarrow proliferation \uparrow apoptosis | [15] |
| Colorectal cancer (continued) | miR-205/YAP1 Axis | SW620, SW480, HT29, and HCT116 | Δ ZEB1-AS1 (which sponges miR-205) \rightarrow \uparrow miR-205 \rightarrow \downarrow YAP1: \downarrow proliferation \uparrow apoptosis | [24] |
| | miR-455-3p/PAK2 axis | SW480, HT29, LS174T, HCT116 and DLD-1 | Δ ZEB1-AS1 (which acts as a molecular sponge) \rightarrow \uparrow miR-455-3p \rightarrow \downarrow PAK2: \downarrow proliferation \downarrow migration \downarrow invasion | [20] |
| | miR-181a-5p/ Wnt/ β -catenin (TCF4) | RKO, Caco2, DLD1, HCT116, LOVO and SW480 | \uparrow miR-181a-5p \rightarrow \downarrow ZEB1-AS1 \rightarrow \downarrow β -catenin & TCF4: \downarrow proliferation \uparrow apoptosis | [38] |
| B-cell acute lymphoblastic leukemia | IL-11/STAT3 | hBMSC-TERT and HS-5 | Δ ZEB1-AS1 \rightarrow \downarrow IL-11 \rightarrow \downarrow STAT3 : \downarrow proliferation | [60] |
| Pancreatic cancer | HIF-1 α /ZEB1 | PANC-1, BXPC-3, AsPC-1, SW1990, and MIAPaCa-2 | Δ HIF-1 α \rightarrow Δ ZEB1-AS1 \rightarrow \downarrow ZEB1: \downarrow proliferation \downarrow migration \downarrow invasion | [23] |
| | miR-505-3p/TRIB2 axis | SW1990, Capan-1 and CFPAC-1 | Δ ZEB1-AS1 (which acts as a molecular sponge) \rightarrow \uparrow miR-505-3p \rightarrow \downarrow TRIB2: \downarrow viability | [64] |

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Table 1 (continued)

| Tumor types | Targets/Regulators and signaling pathways | Cell lines | Functions | Ref |
|---|--|---|--|------|
| Hepatocellular carcinoma | miR-299-3p/E2F1 axis | Huh7, Hep3B, HepG2, MHCC97H and SMMC7721 | ↓ migration ↓ invasion ↑ ZEB1-AS1 (which sponges miR-299-3p) → ↑ E2F1: ↑ proliferation ↑ migration ↑ invasion | [44] |
| | miR-23c | Huh7, SK-HEP-1, SNU-387, JHH-7, and HCCLM3 | Δ ZEB1-AS1 → ↓ miR-23c: ↓ proliferation ↓ invasion | [71] |
| | miR-302b /EGFR/PI3K/AKT axis | PLC, MHCC-97 H, Hep3B, and Huh7 | Δ ZEB1-AS1 (which targets miR-302b) → ↑ miR-302b → ↓ EGFR → ↓ PI3K/AKT pathway: ↓ proliferation ↓ migration ↓ invasion | [17] |
| | miR-365a-3p/E2F2 axis | HepG2, 293 T and HCCLM6 | Δ ZEB1-AS1 (which targets miR-365a-3p) → ↑ miR-365a-3p → ↓ E2F2: ↓ proliferation ↓ viability ↑ cell cycle arrest | [26] |
| Epithelial ovarian cancer | MMP19 | A2780 (including chemoresistant A2780) | ↑ ZEB1-AS1 → ↑ MMP19: ↑ sensitivity to DDP and PTX | [7] |
| Oral Squamous Cell Carcinoma | miR-23a | SCC9, SCC25, TSCCA, HN4, and CAL27 | Δ ZEB1-AS1 (which sponges miR-23a) → ↑ miR-23a: ↓ proliferation ↑ cell cycle arrest ↓ migration ↓ invasion ↓ EMT process | [62] |
| Esophageal squamous cell carcinoma (ESCC)/ Esophageal cancer (EC) | ZEB1 (ESCC) | EC9706, TE1, Eca109, Kyse70 and Kyse450 | Δ ZEB1-AS1 → ↓ ZEB1: ↓ proliferation ↓ invasion | [78] |
| | miR-214/ZEB1 axis (EC) | EC109 | Δ ZEB1-AS1 (which acts as molecular sponge) → ↑ miR-214 → ↓ ZEB1: ↓ proliferation | [75] |
| Prostate cancer | miR-342-3p/CUL4B axis PI3K/AKT/mTOR Signaling pathway | DU145 and LNCaP | Δ ZEB1-AS1 (which acts as molecular sponge) → ↑ miR-342-3p → ↓ CUL4B → ↓ PI3K/AKT/mTOR pathway: ↓ proliferation ↓ migration ↓ invasion | [40] |
| | ZEB1 miR-200c/BMI1 | DU145 and PC3 | Δ ZEB1-AS1 (which recruits MLL1 to the promoter region of ZEB1) → ↓ ZEB1 + ↑ miR-200c → ↓ BMI1: ↓ proliferation ↓ migration | [55] |
| Intrahepatic cholangiocarcinoma | miR-200a/ ZEB1 axis | HuH28, HuCCT1, RBE, CCLP-1 and HCCC-9810 | Δ ZEB1-AS1 (which acts as molecular sponge) → ↑ miR-200a → ↓ ZEB1: ↓ proliferation ↓ migration ↓ invasion ↓ EMT process | [21] |
| Cholangiocarcinoma | AR/ miR-133b/HOXB8 axis | QBC939, CCLP-1, RBE and TFK-1 | Δ ZEB1-AS1 (which is activated by AR) → ↑ miR-133b → ↓ HOXB8: ↓ proliferation ↓ stemness ↓ migration ↓ invasion ↓ EMT process | [20] |
| Thyroid cancer | miR-133a-3p/ LPAR3 /EGFR PI3K/AKT/mTOR signaling pathway | SW579, TPC-1, BCPAP and KAT18 | Δ ZEB1-AS1 (which acts as molecular sponge) → ↑ miR-133a-3p) → ↓ LPAR3 & EGFR → ↓ PI3K/AKT/mTOR: ↓ proliferation ↓ colony formation ↑ apoptosis | [67] |
| Melanoma | - | Primary and metastatic melanoma cell lines | Over expressed ZEB1-AS1 in melanoma, especially metastatic cell lines | [52] |
| | miR-1224-5p | SK-MEL-2, WM35, A375 and SK-MEL-5 | Δ ZEB1-AS1 (which acts as molecular sponge) → ↑ miR-1224-5p ↓ proliferation ↓ migration ↓ invasion | [61] |
| Bladder cancer | miR-200b/FSCN1/ TGF-β1 axis | SW780, BIU, 5637, J82, T24, TCC-SUP, UM-UC3 and RT4 | Δ ZEB1-AS1 (which is overexpressed by TGF-β1) → ↑ miR-200b → ↓ FSCN1: ↓ proliferation ↓ migration | [11] |

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Table 1 (continued)

| Tumor types | Targets/Regulators and signaling pathways | Cell lines | Functions | Ref |
|------------------------------------|---|---|---|------|
| | - | 5637 and SW780 | ↓ invasion ↑ apoptosis Δ ZEB1-AS1 ↓ proliferation ↓ migration | [29] |
| Gastric cancer | miR-149-3p | SGC-7901 and MGC-803 | ↑ apoptosis Δ ZEB1-AS1 → ↑ miR-149-3p: ↓ proliferation ↓ migration ↓ invasion | [39] |
| | - | MKN28, MKN45, BGC823, MGC803, KATOIII, and SGC7901 | Δ ZEB1-AS1: ↓ migration ↓ invasion | [32] |
| | miR-335-5p | SGC7901, HGC27, BGC823, MKN45, MKN28, and AGS | Δ ZEB1-AS1 (which acts as molecular sponge) → ↑ miR-335-5p: ↓ proliferation ↓ migration ↓ invasion | [73] |
| | ZEB1 | NCI-N87, SNU-1, AGS and MKN-45 | Δ ZEB1-AS1 → ↓ ZEB1: ↓ migration ↓ invasion ↓ EMT process | [27] |
| Non-small cell lung cancer (NSCLC) | ID1 | PC9, H1299, H1975, H460, and A-427 | Δ ZEB1-AS1 → ↑ ID1 ↓ proliferation ↓ migration ↑ apoptosis ↑ cell cycle arrest | [22] |
| | miR-409-3p/ZEB1 | | Δ ZEB1-AS1 → ↑ miR-409-3p → ↓ ZEB1: ↓ proliferation ↑ apoptosis | [49] |
| | ZEB1 | A549, H1975, H1299, and SPCA1 (adenocarcinoma) H1703 and SK-MES-1 (squamous carcinoma) | Δ ZEB1-AS1 → ↓ ZEB1: ↓ proliferation ↓ migration ↓ invasion ↑ apoptosis ↑ cell cycle arrest ↓ EMT process | [25] |
| Cervical cancer | ZEB1 | HeLa, C33A and SiHa | Δ ZEB1-AS1 → ↓ ZEB1: ↓ proliferation ↑ apoptosis ↓ invasion ↓ EMT process | [5] |
| Osteosarcoma | miR-200/ZEB1 | HOS and Saos-2 | Δ ZEB1-AS1 (which acts as molecular sponge) → ↑ miR-200 → ↓ ZEB1: ↓ proliferation ↓ migration | [31] |
| | ZEB1 | HOS, U-2 OS, MG-63, and Saos-2 | Δ ZEB1-AS1 → ↓ ZEB1: ↓ proliferation ↓ migration | [30] |

ZEB1-AS1 has been related to the pathologic stage, and its over-expression has been associated with poor prognosis in CRC patients [28]. Other studies in this type of cancer has validated association between ZEB1-AS1 level and clinical stage [72], metastasis [33], histological grade [38], tumor invasion, microvascular invasion and lymph node metastasis [15].

In patients with oral squamous cell carcinoma (OSCC), ZEB1-AS1 expression has been elevated and miR-23a expression level has been lower in cancerous tissues compared to non-cancerous tissues. Notably, ZEB1-AS1 expression levels has been substantially higher in tissues obtained from patients with advanced TNM stages. However, its expression has been similar in different pathological grades [62]. Most notably, expression of ZEB1-AS1 in serum samples of these patients has been decreased after tumor resection. Besides, over-expression of ZEB1-AS1 has been associated with poor survival of patients based on a Kaplan-Meier analysis [62].

In esophageal cancer, up-regulation of ZEB1-AS1 has been verified in both tumor tissues [77] and serum samples [75]. Notably, both peripheral and tumor tissue expressions have been correlated with poor survival. Moreover, its expression in tumoral tissues has been associated with TNM stage, lymph node metastasis [78], tumor grade and depth of invasion [63].

Associations between up-regulation of ZEB1-AS1 and malignant features have also been verified in other types of cancers, including HCC [44], pancreatic cancer [64], prostate cancer [55] and cholangiocarcinoma [20]. Table 3 summarizes these findings.

Diagnostic role of ZEB1-AS1 has been assessed in gastric cancer [39] and colorectal cancer [10] (Table 4). While in the former type of cancer, ZEB1-AS1 can distinguish between tumoral and normal samples with AUC value of 79% [39], in the latter type of cancer, it can separate tumoral and adjacent non-tumoral samples with AUC value of 0.84 [10].

Table 2Impact of ZEB1-AS1 the in carcinogenesis in animal models (Δ : knock-down or deletion, SCID: severe combined immune deficiency).

| Tumor type | Animal models | Results | Ref |
|--|--------------------------|---|------|
| Pancreatic cancer | BALB/c mice | Δ ZEB1-AS1: ↓ tumor growth ↓ metastasis (lung/liver) | [23] |
| Oral Squamous Cell Carcinoma | BALB/c nude mice | Δ ZEB1-AS1: ↓ tumor growth ↓ metastasis | [62] |
| Triple negative breast cancer | Mice | Δ ZEB1-AS1: ↓ tumor size, tumor volume & tumor weight | [36] |
| Hepatocellular carcinoma | Mice | Δ ZEB1-AS1: ↓ micro metastasis | [17] |
| Intrahepatic cholangiocarcinoma | BALB/c nude mice | Δ ZEB1-AS1: ↓ metastasis (liver/lung) ↓ tumor growth | [21] |
| Thyroid cancer | Nude mice | Δ ZEB1-AS1: ↓ tumor growth | [67] |
| Cholangiocarcinoma | BALB/c nude mice | Δ ZEB1-AS1: ↓ tumor growth | [20] |
| Colon adenocarcinoma/colorectal cancer | mice | Δ ZEB1-AS1: ↓ metastasis ↓ tumor growth | [20] |
| | BALB/c athymic nude mice | Δ ZEB1-AS1: ↓ tumor growth | [15] |
| Esophageal Squamous Cell Carcinoma | BALB/c nude mice | Δ ZEB1-AS1: ↓ tumor growth | [78] |
| Bladder cancer | BALB/c nude mice | ↑ ZEB1-AS1: ↑ tumor growth | [11] |
| Glioma | SCID mice | Δ ZEB1-AS1: ↓ metastasis ↓ tumor weight & size | [42] |
| Cervical cancer | BALB/c athymic nude mice | Δ ZEB1-AS1: ↓ tumor growth | [5] |
| Gastric cancer | Nude mice | Δ ZEB1-AS1: ↓ tumor growth ↓ tumor weight | [73] |
| Non-small cell lung cancer (NSCLC) | BALB/c nude mice | Δ ZEB1-AS1: ↓ tumor growth ↓ tumor volume and weight | [22] |
| | BALB/c nude mice | Δ ZEB1-AS1: ↓ tumor growth | [25] |

3. Role of ZEB1-AS1 in non-malignant conditions

3.1. Cell line studies

Different in vitro studies have assessed function of ZEB1-AS1 in non-malignant conditions. For instance, the impact of this lncRNA in the pathophysiology of diabetic lung has been investigate in vitro. Expression of ZEB1-AS1 in lung cells has been found to be decreased after exposure to high concentration of glucose. Moreover, up-regulation of ZEB1-AS1 has led to inhibition of apoptosis in lung cells in association with downregulation of p53. Overexpression of p53 has reduced the effect of ZEB1-AS1 up-regulation on apoptosis of these cells showing that ZEB1-AS1 controls lung cancer cell apoptosis through decreasing expression of p53 [16].

In addition, ZEB1-AS1 has been found to be over-expressed in persistent *Chlamydia trachomatis* infection. This lncRNA participates in the regulation of apoptosis, since its knock-down enhances apoptosis rate in the persistently infected cells. From a mechanistical point of view, ZEB1-AS1 silencing leads to reduction of the Bcl-2/Bax ratio and suppression of the mitochondrial membrane potential leading to cytochrome c release and enhancement of caspase-3 activation. In fact, ZEB1-AS1 acts as a sponge for miR-1224-5p to affect expression of MAP4K4. Moreover, ZEB1-AS1 suppresses apoptosis of *Chlamydia*-infected cells through activation of MAPK/ERK axis [35].

ZEB1-AS1 has also been shown to be upregulated in TGF- β 1-induced RLE-6TN cells in correlation with the level of ZEB1, a master regulator of

EMT. ZEB1-AS1 can enhance fibrogenesis in RLE-6TN cells via modulation of miR-141-3p and activation of ZEB1 in RLE-6TN cells [48]. In addition, ZEB1-AS1 contributes to the pathogenesis of atherosclerosis through modulation of miR-590-5p/ETS1 [3], miR-590-5p/HDAC9 [79] and miR-942/HMGB1 [18] axes. Table 5 summarizes these observations.

The function of ZEB1-AS1 in the pathogenesis of ischemic stroke, diabetic nephropathy and pulmonary fibrosis has been assessed in animal models (Table 6). Expression of ZEB1-AS1 has been shown to be increased in the lungs of BLM-induced rats in correlation with ZEB1 levels. ZEB1-AS1 silencing has relieved BLM-induced fibrogenesis in animal models through suppression of EMT [48]. ZEB1-AS1 has also been shown to aggravate cerebral ischemia/reperfusion injury in animal models through modulation of HMGB1/TLR-4 axis [58].

3.2. Studies in clinical samples

ZEB1-AS1 has been shown to be decreased in the plasma samples of patients with diabetic lung compared with both diabetic persons without this complication and healthy subjects. Notably, downregulated levels of ZEB1-AS1 can distinguish patients with diabetic lung from healthy subjects [16]. On the other hand, ZEB1-AS1 levels have been increased in serum of patients with atherosclerosis relative to normal controls, which has been accompanied by ox-LDL-associated endothelial cell injury in a dose dependent manner [79]. Finally, three independent studies have revealed down-regulation of ZEB1-AS1 in kidney tissues of

Table 3

Dysregulation of ZEB1-AS1 in clinical specimens (ANT: adjacent normal tissue, O-S: overall survival, DFS: disease-free survival, RFS: relapse-free survival, BM: bone marrow, PFS: progression-free survival, GEPIA: Gene Expression Profiling Interactive Analysis, TCGA: the cancer genome atlas, ICGC: The International Cancer Genome Consortium).

| Tumor type | Samples | Expression (tumor vs normal) | Kaplan-Meier Analysis (ZEB1-AS1 dysregulation impact) | Univariate/Multivariate cox regression | Association of ZEB1-AS1 expression with clinicopathologic characteristics | Ref |
|---|--|------------------------------|---|---|--|------|
| Colorectal cancer (CRC)/ colon adenocarcinoma (COAD) | 6 CRC + 6 normal colon tissues | Upregulated | Poor survival rate | Independent prognostic factor | Associated with clinical stage | [28] |
| | TCGA database (429 COAD + 37 normal tissues) | Upregulated | - | - | Associated with advanced clinical stage | [72] |
| | 30 CRC tissues + paired ANT | Upregulated | - | - | - | [19] |
| | TCGA database (647 CRC + 51 normal tissues) | Upregulated | Shorter survival rate | - | Associated with advanced TNM stage | [50] |
| | 20 COAD + paired ANT | Upregulated | - | - | - | [2] |
| | 30 CRC + paired ANT | Upregulated | - | - | Associated with metastasis | [33] |
| | 27 CRC + paired ANT | Upregulated | Poor O-S | - | - | [66] |
| | 50 CRC + paired ANT | Upregulated | - | - | - | [24] |
| | 28 COAD + paired ANT | Upregulated | Poor survival rate | - | Associated with clinical stage, lymph node metastasis and distant metastasis | [20] |
| | 50 CRC + paired ANT | Upregulated | - | - | Associated with TNM stage | [69] |
| | 65 CRC + paired ANT | Upregulated | - | - | Associated with tumor histology grade, N grade, and M grade | [38] |
| | 63 CRC + paired ANT | Upregulated | Shorter O-S | - | Associated with depth of tumor invasion, microvascular invasion and lymph node metastasis | [15] |
| | 108 CRC + paired ANT | Upregulated | Shorter O-S | - | Associated with tumor size, differentiation degree, TNM stage, metastasis, depth of invasion and Dukes' classification | [10] |
| Esophageal Squamous Cell Carcinoma (ESCC)/ Esophageal cancer (EC) | TCGA (174 EC patients) + GSE53622 and GSE53624 (179 ESCC) | Upregulated | Poor survival rate | - | - | [77] |
| | 56 ESCC + paired ANT | Upregulated | Poor survival rate | - | Associated with TNM stage and lymph node metastasis | [78] |
| | Serum of 21 EC patients + 21 healthy controls | Upregulated | - | - | - | [75] |
| | 87 ESCC + paired ANT | Upregulated | Shorter O-S | Independent prognostic factors for O-S | Associated with tumor grade, depth of invasion, and lymph node metastasis | [63] |
| Oral Squamous Cell Carcinoma (OSCC) | 30 OSCC + paired ANT | Upregulated | Shorter O-S | - | Associated with TNM stage | [62] |
| Hepatocellular carcinoma (HCC) | TCGA (319 HCC) + ICGC (230 HCC) | Upregulated | - | - | - | [76] |
| | 60 HCC + paired ANT + GEPIA | Upregulated | Shorter O-S | - | Associated with TNM stage and lymph node metastasis | [44] |
| | 32 HCC + paired ANT | Upregulated | - | - | - | [71] |
| | 90 HCC + paired ANT + TCGA (362 HCC) | Upregulated | Shorter O-S and DFS | Independently associated with the risk of bone metastasis | Associated with metastasis | [41] |
| | 32 HCC + paired ANT | Upregulated | Shorter O-S | - | - | [26] |
| Pancreatic cancer (PC) | 119 PC + paired ANT | Upregulated | Shorter O-S | - | - | [23] |
| | 30 PC + paired ANT | Upregulated | Shorter O-S | - | Associated with advanced clinical stage | [64] |
| | 30 PCa + paired ANT | Upregulated | - | - | - | [40] |
| Prostate cancer (PCa) | 114 paraffin embedded tissues | Upregulated | - | - | Associated with perineural invasion and advanced clinical stage | [55] |
| | 45 TNBC + paired ANT | Upregulated | - | - | - | [36] |
| Triple-negative breast cancer (TNBC) | 45 TNBC + paired ANT | Upregulated | - | - | - | [36] |
| Intrahepatic cholangiocarcinoma (IHCC) | 118 IHCC + 20 paired tissues | Upregulated | Lower O-S and PFS | Independent risk factor for poor O-S and PFS | Associated with microvascular invasion, lymphatic metastasis and advanced TNM stage | [21] |
| Cholangiocarcinoma (CCA) | 54 CCA + paired ANT | Upregulated | Poor O-S | Independent unfavorable prognostic factors for CCA | Associated with lymph node metastasis and TNM stage | [20] |
| Thyroid cancer (TC) | 40 TC + paired ANT | Upregulated | - | - | - | [67] |
| Bladder cancer (BCa) | 60 BCa + 23 paired ANT | Upregulated | Shorter DFS | - | - | [11] |
| | 55 BCa + paired ANT | Upregulated | - | - | Associated with higher histological grade and advanced tumor stage | [29] |
| | 84 GC + 47 gastric dysplasia tissues + 59 healthy gastric mucous | Upregulated | Shorter O-S | Independent unfavorable prognostic factor for GC | Associated with lymph node metastasis, tumor size and TNM stage | [39] |

(continued on next page)

Table 3 (continued)

| Tumor type | Samples | Expression (tumor vs normal) | Kaplan-Meier Analysis (ZEB1-AS1 dysregulation impact) | Univariate/Multivariate cox regression | Association of ZEB1-AS1 expression with clinicopathologic characteristics | Ref |
|---|--|------------------------------|---|---|---|------|
| | 224 GC + paired ANT | Upregulated | Shorter O-S | Independent risk factors for poor prognosis | Associated with TNM stage | [1] |
| | 75 GC + paired ANT | Upregulated | Poor survival rate | Independent prognostic factor | Associated with lymph node metastasis, invasion degree and TNM stage | [32] |
| | 76 GC + paired ANT | Upregulated | Poor survival rate | - | Associated with TNM stage and lymph node metastasis | [73] |
| | 124 GC + 20 paired ANT + 20 gastritis | Upregulated | Shorter O-S | Independent prognostic factor | Associated with lymph node metastasis, clinical stage, histological type and distant metastasis | [27] |
| Non-small cell lung cancer (NSCLC) | 48 NSCLC + 26 pneumonia patients | Upregulated | Shorter O-S | Independent unfavorable prognostic factor | Associated with TNM stage and advanced clinical stage | [22] |
| | 183 NSCLC + 85 paired ANT | Upregulated | Poor survival rate | - | Associated with TNM stage | [68] |
| | 122 NSCLC + paired ANT | Upregulated | Shorter O-S | - | Associated with TNM stage, tumor size and lymph node metastasis | [25] |
| Melanoma | 46 melanoma tissues + paired ANT | Upregulated | Poor survival rate | - | Associated with lymph node metastasis and TNM stage | [61] |
| Glioma | 100 glioma tissues + 16 normal brain tissues | Upregulated | Poor survival rate | - | Associated with tumor size | [42] |
| | 65 glioma tissues + paired ANT | Upregulated | Shorter O-S | Independent predictor of overall survival for glioma patients | Associated with tumor size, KPS and WHO grade | [65] |
| | 82 glioma tissues + paired ANT | Upregulated | Shorter O-S | Independent prognostic factor for glioma patients | Associated with tumor grade | [37] |
| Cervical cancer (CC) | 106 CC + paired ANT | Upregulated | Shorter O-S | - | Associated with FIGO stage and lymph node metastasis | [5] |
| B-cell acute lymphoblastic leukemia (B-ALL) | BM of 30 B-ALL + 30 healthy controls | Upregulated | Poor survival rate | - | - | [60] |
| Osteosarcoma (OS) | 50 OS + paired ANT | Upregulated | Shorter O-S and RFS | - | Associated with tumor size advanced Enneking stage and metastasis | [30] |

patients with diabetic nephropathy [43,53,59]. Table 7 shows human studies on the role of ZEB1-AS1 in non-malignant conditions.

4. Discussion

ZEB1-AS1 is an lncRNA with diverse functions in the pathophysiology of both malignant and non-malignant disorders. It has an important regulatory effect on ZEB1, the master regulator of EMT. Therefore, it can affect pathogenesis of disorders, particularly cancers through induction of EMT and metastasis. Mechanistically, ZEB1-AS1 acts as a sponge for a variety of miRNAs, namely miR-577, miR-335-5p, miR-101, miR-505-3p, miR-455-3p, miR-205, miR-23a, miR-365a-3p, miR-302b, miR-299-3p, miR-133a-3p, miR-200a, miR-200c, miR-342-3p, miR-214, miR-149-3p and miR-1224-5p. These miRNAs can affect carcinogenesis through modulation of expression of cancer-related genes.

Importantly, all conducted studies in the context of cancer have

confirmed an oncogenic role for ZEB1-AS1. This lncRNA can increase cell proliferation, invasiveness, migratory potential and stemness properties. Based on this uniform effect of ZEB1-AS1 in diverse tissues, anti-ZEB1-AS1 strategies can be beneficial for reduction of tumor burden in almost all tissues. Different strategies such as siRNA-mediated gene silencing should be tested in different tissues.

ZEB1-AS1 has a diagnostic value for detection of cancers as revealed in the contexts of gastric cancer [39] and colorectal cancer [10]. Moreover, based on the observed down-regulation of ZEB1-AS1 in serum samples of patients after tumor resection [62], this lncRNA can be used as a marker in follow-up of patients after conduction of anti-cancer therapies.

ZEB1-AS1 has an important role in the induction of chemoresistance. Thus, it is a possible target for combating this phenotype. Combination of anti-ZEB1-AS1 therapies and chemotherapy might be a possible way to reduce chemoresistance.

In addition, up-regulation of ZEB1-AS1 has been linked with several

Table 4

Diagnostic value of ZEB1-AS1 in diseases (ANT: adjacent normal tissue).

| Disease type | Samples | Distinguish between | Area under curve | Sensitivity (%) | Specificity (%) | Ref |
|-------------------------|--|---------------------|------------------|-----------------|-----------------|------|
| Gastric cancer (GC) | 84 GC + 47 gastric dysplasia tissues + 59 healthy gastric mucous | Tumor vs. normal | 0.790 | 82.1 | 79.2 | [39] |
| Colorectal cancer (CRC) | 108 CRC + paired ANT | Tumor vs. normal | 0.846 | 63 | 90 | [10] |

Table 5

Summary of cell line studies on the role of ZEB1-AS1 in non-malignant conditions (Δ : knock-down or deletion, HUVECs: human umbilical vein endothelial cells, ox-LDL: oxidized low-density lipoprotein, HCTAEC: Human carotid artery endothelial cell).

| Disease type | Interactions | Cell line | Function | Ref |
|---------------------------------|---|---|---|------|
| Chlamydia trachomatis infection | miR-1224-5p / MAP4K4 axis | HeLa 229 (infected with C. trachomatis) | \uparrow ZEB1-AS1 \rightarrow sponging of miR-1224-5p \rightarrow \uparrow MAP4K4: \downarrow mitochondria-mediated apoptosis | [35] |
| Pulmonary fibrosis | miR-141-3p / ZEB1 axis | RLE-6TN and 293 T | ZEB1-AS1 acts as a molecular sponge for miR-141-3p \rightarrow \uparrow ZEB1 \rightarrow \uparrow fibrosis | [48] |
| Diabetic nephropathy | miR-216a-5p/BMP7 axis | HK-2 (Treated with high glucose) | \uparrow ZEB1-AS1 (which targets miR-216a-5p) \rightarrow \uparrow BMP7 \downarrow EMT process \downarrow fibrogenesis | [43] |
| | miR-217 / MAFB axis | HK-2 (Treated with high glucose) | \uparrow ZEB1-AS1 (which sponges miR-217) \rightarrow \uparrow MAFB \rightarrow attenuated fibrosis | [53] |
| Atherosclerosis | miR-590-5p / ETS1 Axis TGF- β /Smad Pathway | HUVECs treated with ox-LDL | ZEB1-AS1 sponges miR-590-5p \rightarrow \uparrow ETS1: \uparrow cell injuries & inflammation | [3] |
| | miR-590-5p / HDAC9 axis | HUVECs treated with ox-LDL | Δ ZEB1-AS1 (which sponges miR-590-5p) \rightarrow \uparrow miR-590-5p \rightarrow \downarrow HDAC9 \uparrow proliferation \downarrow apoptosis | [79] |
| | miR-942 / HMGB1 / NOD2 | HCTAEC THP-1 treated with ox-LDL | ZEB1-AS1 sponges miR-942 and upregulates HMGB1 \rightarrow \uparrow cell injuries | [18] |
| | | HUVECs treated with ox-LDL | Δ ZEB1-AS1 (which upregulates NOD2 through recruiting LRPPRC) \downarrow cell injuries \downarrow cell death induced by ox-LDL | [70] |

Table 6

In vivo studies on the role of ZEB1-AS1 in non-malignant conditions.

| Disease type | Animal model | Function | Ref |
|-------------------------|---|---|------|
| Ischemic stroke | Sprague Dawley Rat (ischemia/reperfusion injury models) | \uparrow ZEB1-AS1 \rightarrow \uparrow HMGB1 / TLR-4 axis: \uparrow cognitive impairment \uparrow brain fluid -aggravation of disease | [58] |
| Diabetic nephropathy | C57BL/KsJ mice (diabetic db/db) | \uparrow ZEB1-AS1 \rightarrow reduced renal dysfunction and fibrosis | [53] |
| Pulmonary fibrosis (PF) | Sprague-Dawley rats PF models | Δ ZEB1-AS1 reduces fibrosis | [48] |

malignant features indicating a prognostic role for this lncRNA.

In addition to malignant conditions, ZEB1-AS1 is involved in the pathophysiology of diabetic nephropathy, diabetic lung, atherosclerosis, *Chlamydia trachomatis* infection, pulmonary fibrosis and ischemic stroke through regulation of several miRNA/mRNA axes and signaling pathways.

Taken together, ZEB1-AS1 represents an appropriate target for design of novel therapeutics particularly for cancers where it exerts

Table 7

Summary of human studies on the role of ZEB1-AS1 in non-malignant conditions.

| Disease type | Number of clinical samples | Expression (case vs. control) | Method | Ref |
|---------------------------|---|-------------------------------|---------|------|
| Atherosclerosis (AS) | Blood sample of 30 AS + 30 healthy controls | Upregulated | qRT-PCR | [79] |
| Diabetic lung (DL) | Plasma of 78 DL + 78 + 78 healthy controls | Downregulated in DL | qRT-PCR | [16] |
| Diabetic nephropathy (DN) | 20 DN kidney tissues + 20 healthy controls tissues | Downregulated | qRT-PCR | [43] |
| | 26 DN kidney tissues + 10 normal kidney tissues (from patients with kidney carcinoma) | Downregulated | qRT-PCR | [53] |
| | 8 DN kidney tissues + 8 minimal change disease tissues | Downregulated | qRT-PCR | [59] |

explicit oncogenic roles. Future studies are needed to assess safety and applicability of ZEB1-AS1-targeting strategies in the clinical setting.

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Not applicable.

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CRediT authorship contribution statement

SGF wrote the draft and revised it. MT designed and supervised the study. AA, KBM, BMH and MS collected the data and designed the figures and tables. All the authors read the submitted version and approved it.

Declaration of Competing Interest

The authors declare they have no conflict of interest.

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